Fisheries aquatic sciences aquatic s Fisheries atic sciences aquatic s iences aquatic sciences aquatic s

Spawning biomass of Sardine, *Sardinops sagax*, in waters off South Australia in 2022



Grammer, G. L. and Ivey, A. R.

SARDI Publication No. F2007/000566-12 SARDI Research Report Series No. 1152

> SARDI Aquatics Sciences PO Box 120 Henley Beach SA 5022

> > October 2022

Report to PIRSA Fisheries and Aquaculture





Spawning biomass of Sardine, *Sardinops sagax*, in waters off South Australia in 2022

Report to PIRSA Fisheries and Aquaculture

Grammer, G. L. and Ivey, A. R.

SARDI Publication No. F2007/000566-12 SARDI Research Report Series No. 1152

October 2022

The South Australian Research and Development Institute respects Aboriginal people as the state's first people and nations. We recognise Aboriginal people as traditional owners and occupants of South Australian land and waters. We pay our respects to Aboriginal cultures and to Elders past, present and emerging.

This publication may be cited as:

Grammer, G. L. and Ivey, A.R. (2022). Spawning biomass of Sardine, *Sardinops sagax*, in waters off South Australia in 2022. Report to PIRSA Fisheries and Aquaculture. South Australian Research and Development Institute (Aquatic Sciences), Adelaide. SARDI Publication No. F2007/000566-12. SARDI Research Report Series No. 1152. 27pp.

DISCLAIMER

The authors warrant that they have taken all reasonable care in producing this report. The report has been through the SARDI internal review process, and has been formally approved for release by the Research Director, Aquatic and Livestock Sciences. Although all reasonable efforts have been made to ensure quality, SARDI does not warrant that the information in this report is free from errors or omissions. SARDI and its employees do not warrant or make any representation regarding the use, or results of the use, of the information contained herein as regards to its correctness, accuracy, reliability and currency or otherwise. SARDI and its employees expressly disclaim all liability or responsibility to any person using the information or advice. Use of the information and data contained in this report is at the user's sole risk. If users rely on the information they are responsible for ensuring by independent verification its accuracy, currency or completeness. The SARDI Report Series is an Administrative Report Series which has not been reviewed outside the department and is not considered peer-reviewed literature. Material presented in these Administrative Reports may later be published in formal peer-reviewed scientific literature.

© 2022 SARDI

This work is copyright. Apart from any use as permitted under the *Copyright Act* 1968 (Cth), no part may be reproduced by any process, electronic or otherwise, without the specific written permission of the copyright owner. Neither may information be stored electronically in any form whatsoever without such permission.

Author(s):	Grammer, G. L. and Ivey, A. R.							
Reviewer(s):	urnell, O., Linnane A. (SARDI) and Jones, A. (PIRSA)							
Approved by:	Mayfield, S. Science Leader – Fisheries							
Signed:	Smayfel .							
Date:	31 October 2022							
Distribution:	PIRSA Fisheries and Aquaculture, SARDI Aquatic Sciences, Parliamentary Library, State Library and National Library							
Circulation:	OFFICIAL							

ALL ENQUIRIES

South Australian Research and Development Institute - SARDI Aquatic Sciences 2 Hamra Avenue West Beach SA 5024 PO Box 120 Henley Beach SA 5022 **P:** (08) 8207 5400 **F:** (08) 8207 5415 **E:** <u>pirsa.sardiaquatics@sa.gov.au</u> **W:** <u>http://www.pir.sa.gov.au/research</u>

TABLE OF CONTENTS

EXECU	TIVE SUMMARY	1
1. INT	RODUCTION	2
1.1.	Daily Egg Production Method	2
1.2.	Rationale, objective and approach	2
2. ME	THODS	3
2.1.	Study area and biophysical variables	3
2.2.	Mean daily egg production and spawning area	4
2.3.	Adult reproductive parameters	8
2.4.	Spawning biomass	10
2.5.	Sensitivity analysis	10
3. RE	SULTS	11
3.1.	Distribution and abundance of eggs	11
3.2.	Biophysical variables	12
3.3.	Spawning area	14
3.4.	Mean daily egg production (P_0)	15
3.5.	Adult parameters	17
3.6.	Spawning biomass	18
3.7.	Sensitivity analysis	19
4. DIS	CUSSION	22
4.1.	Egg distribution	22
4.2.	Spawning area and mean daily egg production	22
4.3.	Adult parameters	23
4.4.	Spawning biomass	23
REFER	ENCES	25
APPENI	אוכ	27

LIST OF FIGURES

Figure 1. Map of South Australia showing sites where plankton samples were collected during
2022 and where adult samples were collected with gill-nets between 1998 and 2018
Figure 2. Voronoi nearest neighbour polygons used to estimate the spawning area in 2022. 6
Figure 3. Densities of live Sardine eggs at sites sampled during February and March 2022.
Figure 4. Sea surface temperatures and overlaid with densities of live Sardines eggs at sites
sampled during February and March 202212
Figure 5. Surface concentration of chlorophyll-a overlaid with densities of live Sardines eggs
at sites sampled during February and March 202213
Figure 6. Sampling area and spawning area over the history of DEPM surveys in SA 14
Figure 7. Models fitted to egg densities (eggs.m ⁻²) and egg age (hours) of Sardine cohorts in
2022 (A) and all years combined (1998 to 2022; B)
Figure 8. Estimates of spawning biomass (95% CI) for Sardine in South Australian waters from
1995 to 2022 using the log-linear egg production model and all-years data for all
parameters, except for spawning area (A)19
Figure 9. Sensitivity plots showing effects of variability in adult parameters and egg production
on estimates of spawning biomass21

LIST OF TABLES

Table 1. Total area surveyed and spawning area (A) estimated in 2022	
Table 2. Mean daily egg production (P_0) and instantaneous daily mortality ((Z) for all data
collected from 1998 to 2022	15
Table 3. Mean daily egg production (P_0) and instantaneous daily mortality (Z)	for data solely
collected in 2022	15
Table 4. Parameter estimates used in the calculations of spawning biomass	18

ACKNOWLEDGEMENTS

We thank the master and crew of the *RV Ngerin*, Zoran Koncurat, Slavko Kolega, Dean Eyre, Jonathan Western, Nick Jobson, and Gaynor Crosby for their assistance during the research surveys. Leonardo Durante, Callum White, Andrew Hogg, Chris Presser, and Fred Bailleul provided technical assistance during the surveys. We thank Associate Professor Tim Ward for providing his in-depth knowledge and countless contributions to the research and management of the South Australian Sardine Fishery over many years. Drs Owen Burnell and Adrian Linnane reviewed and provided comments on a draft of the report. The report was approved for publication by Dr Stephen Mayfield.

EXECUTIVE SUMMARY

This report provides an estimate of the spawning biomass of Sardine, *Sardinops sagax*, in waters off South Australia during 2022. The estimate of spawning biomass obtained using the Daily Egg Production Method (DEPM) is the key performance indicator for determining the status of the southern stock of Australian Sardine.

An ichthyoplankton survey was conducted during February/March 2022. The survey area was expanded in 2020 to include 40 new sites south-east of Kangaroo Island, and this was continued in 2022. The total survey area was 129,982 km². Live Sardine eggs were collected at 218 of 381 (57.2%) sites. The total spawning area (*A*) in 2022 was 76,842 km². Removing the additional sites added in 2020 and 2022 for comparison to previous years, reduced the spawning area to 70,576 km² (third highest on record). Mean daily egg production (P_0 , 95% CI) was 83.6 (75.5–92.7) eggs.day⁻¹.m⁻². P_0 was estimated using the linear version of the exponential mortality model and all data combined from 1998 to 2022.

Estimates of adult parameters (95% CI) calculated from all data obtained between 1998 and 2018 were: sex ratio (*R*): 0.55 (0.52–0.58); spawning fraction (*S*): 0.108 (0.100–0.119); and relative fecundity (*F'*): 305.4 (304.2–306.6) eggs.g⁻¹.

Sensitivity analyses showed the effects of inter-annual variability in parameters (i.e. P_0 , R, S and F') on the estimate of spawning biomass for 2022 and demonstrated the benefits of using parameter estimates obtained from historical data to estimate spawning biomass.

The estimate of spawning biomass (95% CI) of Sardine for 2022 was 355,075 (299,956–410,193) t, which is the second highest on record after 2020 (1995 to 2022) and above the upper target reference point of 190,000 t set in the harvest strategy for the South Australian Sardine Fishery. On this basis, the southern stock of Australian Sardine is classified as **Sustainable**. This classification is consistent with the findings of the spawning biomass report for 2020, the stock assessment report for 2021 and the most recent report in the Status of Australian Fish Stocks.

Keywords: Sardine, Spawning Biomass, South Australia, Daily Egg Production Method.

1. INTRODUCTION

1.1. Daily Egg Production Method

The Daily Egg Production Method (DEPM) was developed for stock assessment of the Northern Anchovy (*Engraulis mordax*; Parker 1980, Lasker 1985), and has been applied to more than 20 species of small to medium-sized pelagic fishes (e.g. Stratoudakis et al. 2006, Dimmlich et al. 2009, Neira et al. 2009, Grammer et al. 2022). The method is widely used in coastal fisheries because it is often the most practical option available for assessment of pelagic species (Ward et al. 1998).

The DEPM relies on the premise that spawning biomass can be calculated by dividing the mean number of pelagic eggs produced per day throughout the spawning area (i.e. total daily egg production) by the mean number of eggs produced per unit mass of adult fish (i.e. mean daily fecundity, Parker 1980, Lasker 1985). Total daily egg production is the product of mean daily egg production (P_0) and total spawning area (A). Mean daily fecundity is the product of mean sex ratio (by weight, R), mean spawning fraction (proportion of mature females spawning each day/night, S) and mean relative fecundity (number of eggs produced per gram of total female weight (\hat{F}/W), F). Spawning biomass (SB) is calculated according to the equation:

$$SB = P_0 * A/(R * S * F')$$
 Equation 1

The DEPM, as applied to Sardine off South Australia, underwent a comprehensive review in 2021 (Ward et al. 2021). The review by Ward et al. (2021) reanalysed data collected between 1995 and 2019 for South Australian Sardine and identified several ways to increase the precision of estimates of spawning biomass: 1) increase the precision of totally daily egg production (P_0 *A) by using the estimate of P_0 obtained from all historical data rather than annual estimates of P_0 ; 2) continue to use the log-linear model to estimate P_0 for the southern stock of Australian Sardine; 3) increase the precision of mean daily fecundity (R*S*F') by using the estimates of sex ratio (R), spawning fraction (S) and relative fecundity (F') obtained from all historical data rather than annual estimates; 4) combine batch fecundity (F) and female weight (W) into a single parameter: relative fecundity ($F' = \hat{F}/W$). These recommendations have been implemented for this report to ensure that estimates of spawning biomass are, and continue to be, as accurate and precise as possible.

1.2. Rationale, objective and approach

The DEPM has been used to estimate the spawning biomass of Sardine in South Australian waters since 1995 (Ward et al. 1998, 2011, 2020, 2021). The estimate of spawning biomass

obtained using the DEPM is the key performance indicator for determining the status of the southern stock of Australian Sardine (PIRSA 2014). The objective of this report is to estimate the spawning biomass of Sardine in waters off South Australia in 2022. Annual estimates of P_0 and A were obtained from an ichthyoplankton survey conducted in 2022. Adult parameters were derived from all adult samples collected off South Australia between 1998 and 2018; an estimate of P_0 was also obtained from all historical data. Sensitivity analyses were undertaken to evaluate the effects of variability in estimates of individual parameters on the uncertainty associated with the estimate of spawning biomass for 2022.

2. METHODS

2.1. Study area and biophysical variables

2.1.1. Study area

An ichthyoplankton survey was conducted from the *RV Ngerin* in shelf and gulf waters of South Australia during February and March 2022 (Fig. 1). Plankton samples were collected at a total of 381 sites on 38 transects between Kingston and the Head of Bight (Fig. 1). The 381 sites sampled in 2022 included 40 additional sites southeast of the traditional survey area (Fig. 1). These additional sites were added in 2020 to the south-eastern end of the survey and east of Kangaroo Island in response to the expansion of the fishery into this region and recent observations that Sardine eggs had become more common in the area (Ward et al. 2020; Fig. 1).

An adaptive approach to egg sampling has been applied since 2014 to ensure that each survey covers as much of the spawning area as possible. Adaptive sampling was implemented in response to the incomplete coverage of the spawning area in 2013 (see Grammer et al. 2021). Under the protocol, extra samples are taken at sites located outside the area covered by the pre-determined sampling sites (Fig. 1). Decisions about whether (or not) to take extra samples are based on the presence/absence of eggs in samples taken using the Continuous Underway Fish Egg Sampler (CUFES) at sites located on the seaward end of transects. Sampling at additional sites continues until Sardine eggs are not present in the CUFES samples. In 2022, two extra sites were sampled on the seaward ends of two transects.

2.1.2. Water temperature and primary production

At each sampling site (Fig. 1), a *Sea-Bird* Conductivity-Temperature-Depth (CTD) recorder fitted with a fluorometer was lowered to a depth of 70 m, or to 10 m from the bottom in waters less than 80 m deep. Estimates of water temperature and fluorescence at the surface were

extracted from each CTD profile. At sites where water temperature was not recorded (due to technical difficulties), the average temperature of the adjacent stations was applied. Fluorescence is an indicator of primary production and gives an un-calibrated measure of chlorophyll-*a* concentration (µg.L⁻¹). Spatial plots of sea surface temperature (SST) and chlorophyll-*a* concentration were prepared using minimum curvature algorithms in Surfer[®] (Ver. 8).

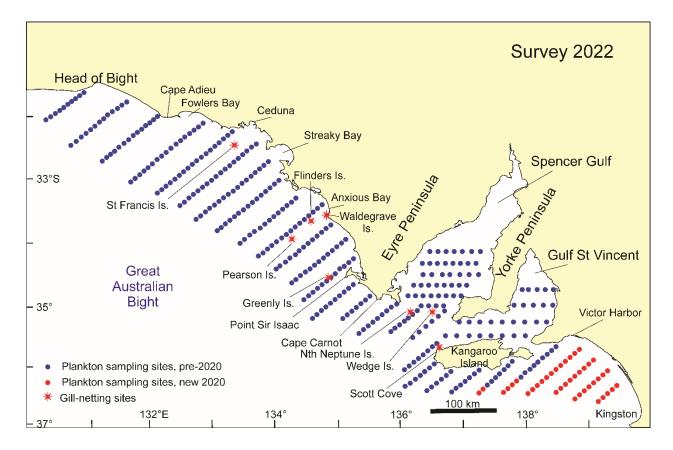


Figure 1. Map of South Australia showing sites where plankton samples were collected during 2022 and where adult samples were collected with gill-nets between 1998 and 2018. Red dots: new sites added to the survey are since 2020.

2.2. Mean daily egg production and spawning area

2.2.1. Plankton sampling

Plankton samples were collected at each site using paired Californian Vertical Egg Tow (CalVET) plankton nets. Each CalVET net had an internal diameter of 0.3 m, length of 1.8 m, 330 μ m mesh and plastic removable cod-ends. During each tow the CalVET nets were deployed to a depth of 70 m or to 10 m of the seabed at depths <80 m. The nets were retrieved vertically at a speed of ~1 m.s⁻¹. *General Oceanics* 2030 flow-meters and factory calibration

coefficients were used to estimate the distance travelled by the net during each tow. Where there was a discrepancy of more than 5% between flow-meters, the relationship between wire length and flow-meter units was used to determine which was correct and that value was used for both nets. Upon retrieval of the nets, the samples from the two cod-ends were rinsed with seawater, combined into a 1L container, and stored in 5% buffered formaldehyde and seawater solution.

2.2.2. Laboratory analysis

Sardine eggs and larvae were identified in each plankton sample using published descriptions (Neira et al. 1998, White and Fletcher 1998). Eggs were staged based on descriptions in White and Fletcher (1998). Total counts of eggs of each developmental stage in each sample were recorded. Eggs in the first and last stages were excluded from the statistical analyses as they were under- and over-represented in samples, respectively (Ward et al. 2018a).

2.2.3. Egg ageing and treatment of zero count egg samples

The development rate of Sardine eggs is dependent on ambient water temperature (Picquelle and Stauffer 1985, Pauly and Pullin 1988). Based on the temperature data from the CTD, egg samples were allocated to one of three temperature bins that covered the range of temperatures encountered during surveys (14–18°C, 18–22°C, and 22–26°C). The temperature bins were comparable to those used by Le Clus and Malan (1995) to describe the developmental rates of Sardine eggs. These published development rates were used to assign a mean age to each egg in each sample (Ward et al. 2018a).

After each egg was assigned an age, the eggs in each sample were grouped into daily cohorts. This was done because a sample usually included eggs spawned on more than one night. The total number of eggs in each daily cohort was calculated by summing the number of eggs of each stage assigned to a spawning day (i.e. day 0, day 1, day 2). The age of a daily cohort was calculated from the average age of each stage within the daily cohort, weighted by the number of eggs in each stage.

Samples with eggs could contain several possible combinations of daily cohorts depending on water temperature, spawning time (peak around 2:00 am) and sampling time. Zero counts were allocated for daily cohorts where the cohort was expected to be present but was not found within the sample (Ward et al. 2018a). Samples with no eggs were excluded from the analyses and not considered part of the spawning area.

5

2.2.4. Egg density

The number of eggs of each day class under one square metre of water (P_t) was estimated at each site according to Equation 2:

$$P_t = \frac{C.D}{V}$$
 Equation 2

Where *C* is the number of eggs of each age in each sample, *V* is the volume filtered (m^3), and *D* is the depth (m) to which the net was deployed (Smith and Richardson 1977). Plots of egg distribution and abundance were prepared using Surfer[®] (Ver. 8).

2.2.5. Spawning area (A)

The spawning area (*A*, Lasker 1985, Somarakis et al. 2004) was estimated using the Voronoi natural neighbour method (Watson 1981). The survey area was divided into a series of contiguous polygons approximately centered on each site using the 'deldir' package in the statistical program 'R (Fig. 2; R Development Core Team 2022, Turner 2016). The area represented by each site (km²) was calculated. *A* was defined as the total area of the polygons where live Sardine eggs were present in the plankton sample (see Fletcher et al. 1996).

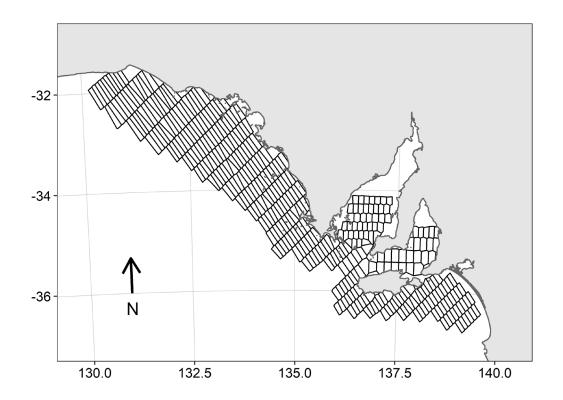


Figure 2. Voronoi nearest neighbour polygons used to estimate the total spawning area in 2022.

2.2.6. Mean daily egg production (P_0) and egg mortality (Z)

The underlying model used to calculate P_0 was the exponential egg mortality model (Equation 3) with a bias correction factor (Equation 4, the 'log-linear model'). The linear version of the exponential egg mortality model is:

$$\ln P_b = \ln (P_{i,t} + 1) - Zt, \qquad \text{Equation 3}$$

where $P_{i,t}$ is the density of eggs of age t at site i and Z is the instantaneous rate of egg mortality.

Estimates of P_b obtained using the linear version of the exponential mortality model have a negative bias, therefore a bias correction factor was applied following the equation of Picquelle and Stauffer (1985):

$$P_0 = e^{\left(\ln P_b + \sigma^2/2\right)} - 1$$
 Equation 4

where, σ^2 is the variance of the estimate of biased mean daily egg production (P_b).

A general linear model (GLM) with a negative binomial error structure (NB1) was also used to estimate P_0 (Equation 5):

$$E[P_0] = g^{-1}(-zt + \varepsilon)$$
 Equation 5

where $E[P_0]$ is the expected value of P_0 , g^{-1} is the inverse-link function, *zt* is the instantaneous rate of daily egg mortality at age *t*, and ε is the error term. The negative binomial error structure used is considered suitable for over-dispersed data, such as egg density by age (e.g. Ward et al. 2011, 2018, 2021). For NB1, variance increased linearly with the mean ($\sigma = \mu^*(1 + \mu + \phi)$), where μ is the model estimate, σ is the model variance and ϕ is the over-dispersion parameter. The GLM used a log-link function (Wood 2006) and was fit using the glmmTMB 'R' package (Brooks et al., 2017).

Following the recommendations of Ward et al. (2021), the value of P_0 from the log-linear model was used to estimate spawning biomass for Sardine. P_0 was calculated using data collected solely in 2022, as well as with data from all years (combined) between 1998 and 2022. The all-years estimate of P_0 is considered more robust than the individual year estimate of P_0 , because sampling error within a year is greater than inter-annual variability of egg density and egg production (Ward et al. 2021).

2.3. Adult reproductive parameters

Adult parameters used to estimate spawning biomass were derived from all adult samples of Sardine collected for DEPM surveys off South Australia between 1998 and 2018 (see Ward et al. 2021)

2.3.1. Sampling methods

From 1998 to 2018, samples of mature Sardine were collected from sites located in the eastern Great Australian Bight, southern Spencer Gulf and Investigator Strait using a gill-net (Fig. 1). In the late afternoon, a dual frequency echo sounder (60 and 180 KHz) was used to search areas where schools of adult Sardine were known to aggregate. A gillnet comprised of three panels, each with a different multi-filament nylon mesh size (*Double Diamond*: 210/4 ply meshes 25, 28 and 32 mm) was deployed from the port side of the *RV Ngerin* at protected locations where schools were encountered. Surface and sub-surface lights (150 W) were illuminated near the net after it was set. Net soak times varied from 15 minutes to 3 hours depending on the number of fish caught.

After the net was retrieved, fish were removed and dissected immediately. All Sardine collected were counted and sexed. Mature males and immature fish were frozen. Mature females were fixed in 10% buffered formaldehyde seawater solution.

2.3.2. Female weight (W) and Male weight

Mature females from each sample were removed from the formalin solution and weighed (\pm 0.01 g). Fixation in formalin has a negligible effect on fish weight (Lasker 1985). The mean weight of mature females in the population was calculated from the average of sample means weighted by proportional sample size:

$$W = \boxed{\overline{W_i} * \frac{n_i}{N}}$$
Equation 5

where, $\overline{W_i}$ is the mean female weight of each sample *i*; *n* is the number of fish in each sample and *N* is the total number of fish collected in all samples.

Mature males in each sample were thawed and weighed (± 0.01 g).

2.3.3. Sex ratio (R)

The mean sex ratio of mature individuals in the population was calculated from the average of sample means weighted by sample size:

$$R = \left[\overline{R_i} * \frac{n_i}{N}\right]$$

where, *n* is the number of fish in each sample, *N* is the total number of fish collected in all samples and $\overline{R_i}$ is the mean sex ratio of each sample calculated from the equation:

$$\overline{R_i} = \frac{F}{(F+M)}$$
 Equation 7

where, F and M are the respective total weights of mature females and males in each sample i.

2.3.4. Spawning fraction (S)

Ovaries of mature females were sectioned and stained with haematoxylin and eosin. Several sections from each ovary were examined to determine the presence/absence of post-ovulatory follicles (POFs). POFs were aged according to the criteria developed by Hunter and Goldberg (1980) and Hunter and Macewicz (1985). The spawning fraction of each sample was estimated as the mean proportion of females with hydrated oocytes plus day-0 POFs (*d*0) (assumed to be spawning or have spawned on the night of capture), day-1 POFs (*d*1) (assumed to have spawned the previous night) and day-2 POFs (*d*2) (assumed to have spawned the previous night) and day-2 POFs (*d*2) (assumed to have spawned two nights prior). The mean spawning fraction of the population was then calculated from the average of sample means weighted by proportional sample size:

$$S = \left[\overline{S_i} * \frac{n_i}{N}\right]$$
 Equation 8

where, *n* is the number of fish in each sample, *N* is the total number of fish collected in all samples and $\overline{S_i}$ is the mean spawning fraction of each sample calculated from the equation:

$$\overline{S_i} = \frac{\left[(d0 + d1 + d2POFs)/3 \right]}{n_i}$$
 Equation 9

where, d0, d1 and d2 POFs are the number of mature females with POFs in each sample and n_i is the total number of females within a sample.

2.3.5. Batch fecundity (F)

Batch fecundity (F) was estimated from ovaries containing hydrated oocytes using the methods of Hunter and Macewicz (1985). Both ovaries were weighed and the number of

hydrated oocytes in three weighed ovarian sub-sections counted. The total batch fecundity for each female was calculated by multiplying the mean number of oocytes per gram of ovary segment by the total weight of the ovaries. Methods to estimate the batch fecundity for mature females without hydrated ovaries (\hat{F}) followed those outlined in Ward et al. (2021).

Relative Fecundity (F') was calculated by using the linear relationship of batch fecundity determined from all years data (1998-2018) to estimate F and then dividing by the mean weight of all mature females collected (W).

2.4. Spawning biomass

Spawning biomass was calculated using the all-years estimate of P_0 (1998 to 2022) obtained from the log-linear model, spawning area (*A*) in 2022 and estimates of *R*, *S* and *F*' obtained from adult samples collected between 1998 and 2018.

The reliability of model fits, 95% confidence intervals (CIs) and coefficients of variation (CVs) for P_0 were estimated using bootstrap resampling methods with 10,000 iterations. Coefficients of variation and CIs for *R*, *S* and *F*', were calculated from the all-years adult data. A ratio estimator was used calculate the variance for *S*, *R*, and *F*' (Rice 1995). The variance for the spawning biomass estimates were calculated by summing the squared CVs for each parameter and multiplying by the square of the estimate of spawning biomass (Parker 1985). Uncertainty estimates presented for all parameters are 95% CIs. Data analyses were done in the R programming environment (R Core Team 2022).

2.5. Sensitivity analysis

Sensitivity analyses were conducted to assess the effects of variations in the range of values obtained for each parameter in each year between 1998 and 2022 on the estimate of spawning biomass for 2022.

3. RESULTS

3.1. Distribution and abundance of eggs

A total of 5,252 live Sardine eggs were collected at 218 of 381 (57.2%) sites on 38 transects between the Kingston and Head of Bight between February and March 2022 (Fig. 3). Sites with the highest egg densities were located in the mouth of Spencer Gulf, southeast of Kangaroo Island and on the shelf in the central Great Australian Bight (GAB).

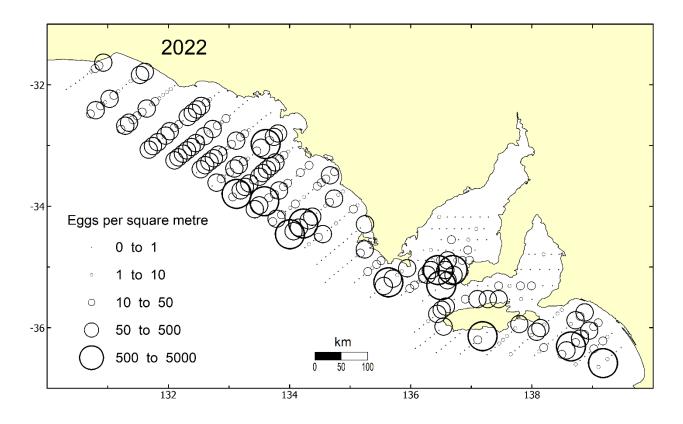


Figure 3. Densities of live Sardine eggs at sites sampled during February and March 2022.

3.2. Biophysical variables

3.2.1. Sea surface temperature

Sea surface temperatures (SSTs) ranged from 15.2 to 22.6°C (Fig. 4) between February and March 2022. High SSTs (>20°C) were recorded in Spencer Gulf, Gulf St. Vincent and throughout the central GAB. Cooler, upwelled water (<19°C) was widespread throughout the eastern GAB and to the east and southeast of Kangaroo Island.

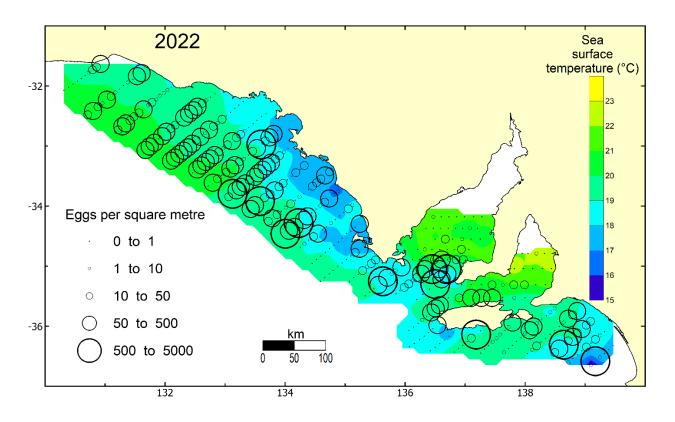


Figure 4. Sea surface temperatures and overlaid with densities of live Sardines eggs at sites sampled during February and March 2022.

3.2.2. Fluorescence

Surface chlorophyll-*a* concentration at each site ranged between 0.02 and 6.70 μ g.L⁻¹ (Fig. 5) between February and March 2022. The highest values were recorded in the Gulfs and in the southeast of the survey. The remainder of coastal and shelf waters mainly had chlorophyll-*a* concentrations <1.0 μ g.L⁻¹.

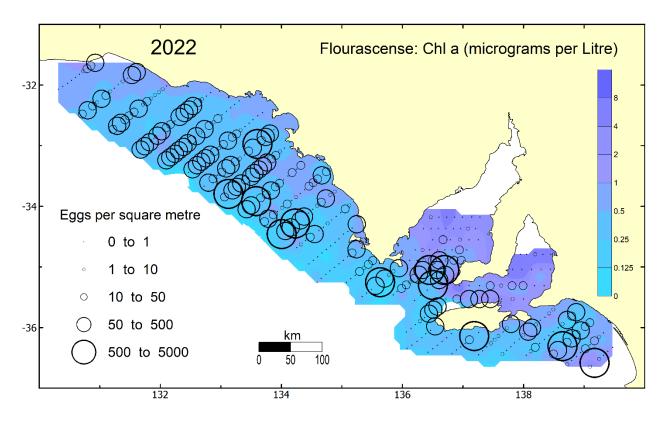


Figure 5. Surface concentration of chlorophyll-*a* overlaid with densities of live Sardines eggs at sites sampled during February and March 2022.

3.3. Spawning area

The estimated spawning area was 76,842 km² and comprised 59.1% of the total area sampled (129,982 km², Table 1, Fig. 6). If the additional transects sampled in 2022 were not included, the spawning area would have been 70,576 km² (Figs. 1 and 6, Table 1).

Table 1. Total	area surveyed and	spawning area (A) estimated in 2022.

	Area sampled (km ²)	Spawning area, <i>A</i> (km²)	Spawning area percentage
Pre-2020 survey	118,735	70,576	59.4
With new stations	129,982	76,842	59.1

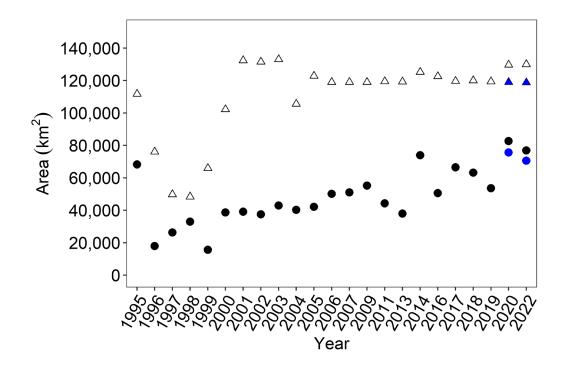


Figure 6. Sampling area (open triangles) and spawning area (closed circles) over the history of DEPM surveys in South Australia. For comparison: blue points for 2020 and 2022 exclude additional sites added since 2020.

3.4. Mean daily egg production (Po)

The estimate of P_0 (95% CI) obtained by fitting the log-linear model (Eq. 3) to all data from 1998 to 2022 was 83.6 (75.5–92.7) eggs.day⁻¹.m⁻² (Table 2, Fig. 7). The estimate of P_0 obtained by fitting the GLM NB1 (Eq. 5) to all data from 1998 to 2022 was 97.1 (82.7–115.9) eggs.day⁻¹.m⁻² (Table 2, Fig. 7).

Table 2. Mean daily egg production (P_0) and instantaneous daily mortality (Z) estimated by fitting the log-linear model and GLM NB1 to all data collected from 1998 to 2022.

Model fit	<i>P₀</i> (eggs·day ⁻¹ ·m ⁻²) (95% CI)	Z				
Log-linear model	83.6 (75.5–92.7)	0.52 (0.44–0.59)				
GLM, Negative Binomial	97.1 (82.7–115.9)	0.35 (0.29–0.41)				

The estimate of P_0 (95% CI) obtained by fitting the log-linear model (Eq. 3) to data solely obtained in 2022 was 97.6 (66.4–141.3) eggs.day⁻¹.m⁻² (Fig. 7, Table 3). The estimate of P_0 obtained by fitting the GLM NB1 (Eq. 5) to data solely obtained in 2022 was 92.4 (66.4–125.7) eggs.day⁻¹.m⁻² (Fig. 7, Table 3). Estimates of P_0 for individual years ranged from 39.0 in 2013 to 145.3 in 2004 (Appendix 1).

Table 3. Mean daily egg production (P_0) and instantaneous daily mortality (Z) estimated by fitting using the log-linear model and GLM NB1 to data solely collected in 2022.

Model fit	<i>P₀</i> (eggs·day ⁻¹ ·m ⁻²) (95% CI)	Z				
Log-linear model	97.6 (66.4–141.3)	0.53 (0.18–0.85)				
GLM, Negative Binomial	92.4 (66.4–125.7)	0.33 (0.08–0.58)				

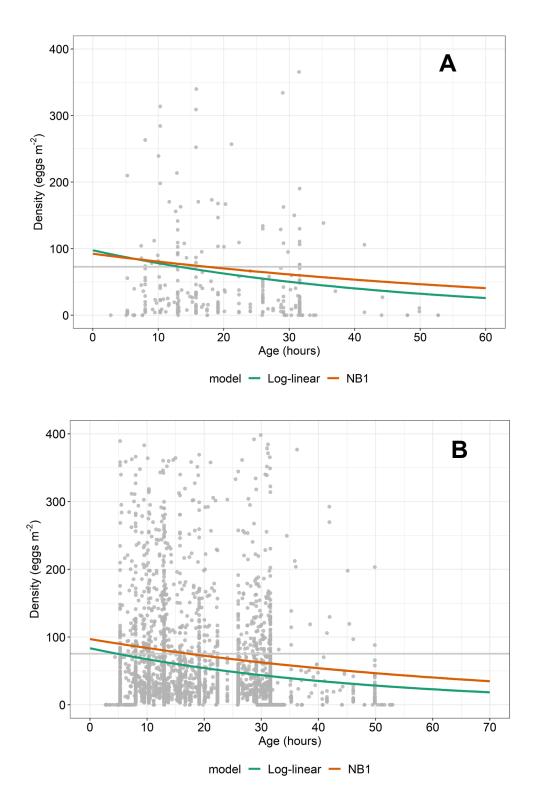


Figure 7. Models fitted to egg densities (eggs.m⁻²) and egg age (hours) of Sardine cohorts in 2022 (**A**) and all years combined (1998 to 2022; **B**). Grey horizontal line: mean egg density.

3.5. Adult parameters

3.5.1. Mean female weight

The mean weight of mature females (W, 95% CI) estimated from 16,995 fish (255 samples) collected between 1998 and 2018 was 58.4 g (23.1–93.7) (Ward et al. 2021). Estimates of W for individual years ranged between 46.5 g in 1998 and 78.7 g in 2004 (Appendix 1).

3.5.2. Sex ratio

The mean sex ratio by weight (R, 95% CI) calculated from all fish collected between 1998 and 2018 was 0.55 (0.52–0.58) (Ward et al. 2021, Table 4). Estimates of R for individual years ranged from 0.36 in 2009 to 0.70 in 2018 (Appendix 1).

3.5.3. Batch fecundity

Between 1998 and 2018, 1,099 females with hydrated oocytes were collected (Ward et al. 2021). The fecundity-weight relationship estimated from these samples was: Batch Fecundity = 335 × Gonad Free Female Weight – 797 ($R^2 = 0.53$) (Ward et al. 2021). Mean gonad free female weight between 1998 and 2018 was 55.5 g and ranged between 43.2 and 75.0 g (Appendix 1). Overall mean batch fecundity (\hat{F} , 95% CI) was 17,835 (3,790–31,880) oocytes (Appendix 1).

The overall estimate of *F*' was 305.4 (95% CI: 304.2–306.6) eggs.g⁻¹ (Ward et al. 2021, Table 4). Estimates of *F*' for individual years ranged from 295.9 eggs.g⁻¹ in 2000 to 312.9 eggs.g⁻¹ in 2011 (Appendix 1).

3.5.4. Spawning fraction

The spawning fraction (S, 95% CI) calculated from all data collected between 1998 and 2018 was 0.108 (0.100–0.119) (Ward et al 2021, Table 4). A total of 16,334 ovaries were examined; 2,578 had day-0 POFs or hydrated oocytes, 1,540 had day-1 POFs and 1,046 day-2 POFs (Ward et al. 2021). Estimates of S for individual years ranged from 0.041 in 2014 to 0.179 in 2001 (Appendix 1).

Table 4. Parameter estimates used in the calculations of spawning biomass. Estimates of adult parameters (R, S, F') are calculated from all historical data from 1998 to 2018 (Ward et al. 2021).

Parameter	All Years	95% CI	CV	Range (among years)
Egg Production (<i>P</i> ₀ , eggs.day ⁻¹ .m ⁻²)	83.6	75.5–92.7	0.052	39.0–145.3
Sex Ratio (<i>R</i>)	0.55	0.52-0.58	0.026	0.36–0.70
Spawning Fraction (S)	0.108	0.100–0.119	0.053	0.041–0.179
Relative Fecundity (<i>F</i> ', eggs.g ⁻¹)	305.4	304.2–306.6	0.002	295.9–312.9
Spawning Area (A, km²)	-	-	-	15,637–82,627

3.6. Spawning biomass

The estimate of spawning biomass (95% CI) calculated using the estimate of *A* obtained from the survey conducted in 2022, and the all-years estimates of P_0 (log-linear model), *S*, *R*, and *F*' was 355,075 (299,956–410,193) t (Fig. 8). The estimate calculated using the value of *A* without the additional stations added since 2020 was 326,119 t.

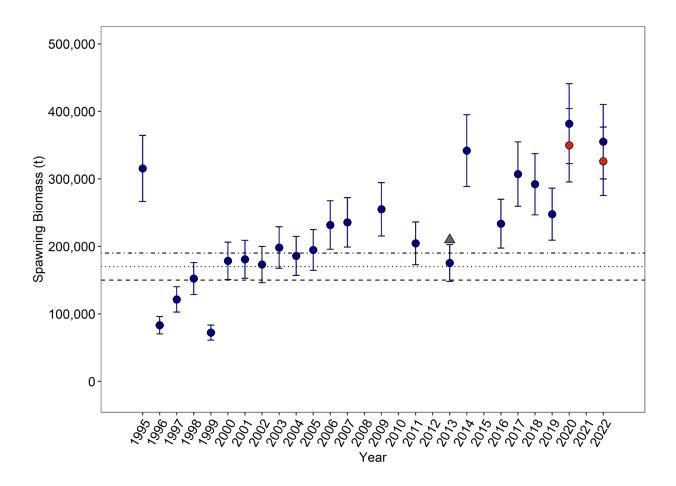


Figure 8. Estimates of spawning biomass (95% CI) for Sardine in South Australian waters from 1995 to 2022 using the log-linear egg production model and all-years data for all parameters, except for spawning area (*A*). The red circles for 2020 and 2022 are the estimate of spawning biomass obtained using estimate of *A* without the additional stations added since 2020. The grey triangle for 2013 (when the survey did not cover the entire spawning area) is the estimate of spawning biomass using the mean *A* from 2002 to 2011 (45,406 km²). The horizontal lines indicate the 150,000 t (dash), 170,000 t (dotted) and 190,000 t (dash/dot) reference points in the harvest strategy (PIRSA 2014).

3.7. Sensitivity analysis

The sensitivity analysis shows the effects of inter-annual variability in parameters (i.e. P_0 , R, S and F') on the estimate of spawning biomass for 2022 (Table 4, Fig. 9, Appendix 1).

The high level of inter-annual variability in the estimates of P_0 reflects the high level of statistical uncertainty associated with annual estimates of this parameter (Fig. 9, Appendix 1) (see Ward et al. 2021). This range of variability had a strong influence on the estimate of spawning biomass (i.e. 166,000 to 617,000 t; Fig. 9). The estimate of spawning biomass obtained using the P_0 estimated from 2022 survey was higher than the estimate obtained using all-years data. Both estimates of P_0 were within the range of values obtained in individual years between 1998 and 2020 (Fig. 9, Table 4).

The estimates of *R* obtained in individual years were variable (Table 4; Appendix 1). The extreme values (i.e. 0.36 and 0.70) are likely due to the limitations of the adult sampling program rather than the relative abundance of sexes in the population (see Ward et al. 2021). The variations in *R* had a large influence on the estimate of spawning biomass for 2022 (i.e. 277,000 to 547,000), which demonstrates the benefits of using all-years data to estimate this parameter.

The estimates of spawning fraction (*S*) obtained in individual years were also highly variable (i.e. ranging from 0.041 to 0.179). Inter-annual variations in *S* are more likely to reflect the limitations of the adult sampling program than differences in the spawning rates occurring in the population (see Ward et al. 2021). Inter-annual variability is *S* had the strongest influence of all of the parameters on the estimate of spawning biomass (i.e. 213,000 to 926,000 t), which demonstrates the benefits of using all-years data to estimate this parameter.

The estimate of relative fecundity (F') was similar among years and inter-annual variation in this combined parameter had a much smaller effect on spawning biomass (i.e. 347,000 to 367,000 t, Fig. 9). Inter-annual variations in W and F between 1998 and 2018 were large and have a strong influence on the estimate of spawning biomass (Appendix 1; see Ward et al. 2021). Estimating F' as a single parameter, as done in the original formulation of the DEPM (Parker, 1980), from all historical data greatly increases the precision of estimates of spawning biomass for Sardine (Ward et al. 2021).

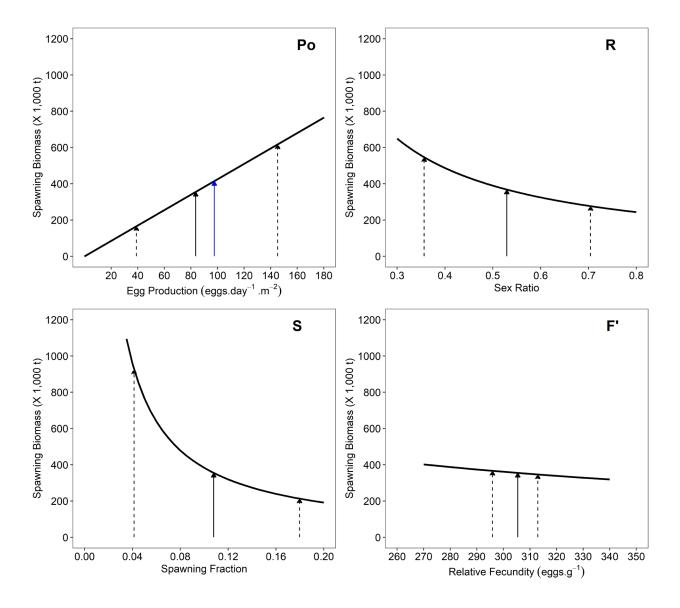


Figure 9. Sensitivity plots showing effects of variability in adult parameters and egg production on estimates of spawning biomass. Solid black arrows: parameter estimates for all years combined; Dashed arrows: range of values recorded between 1998 and 2018; Blue arrow: P_0 estimate using only data collected in 2022.

4. DISCUSSION

4.1. Egg distribution

The distribution of Sardine eggs off South Australia in 2022 was largely comparable to previous years. Similar to 2020, Sardine eggs occurred in almost all of the areas where large numbers of eggs have been collected historically (Grammer et al. 2021, Ward et al. 2021). In contrast, in most previous years there were some areas where few eggs were collected, such as off the southern Eyre Peninsula in 2019 and the central GAB in 2018 (Ward et al 2019, 2020). Since 2020, 40 additional sites were sampled to the south-east of the traditional survey area. This additional sampling was undertaken in response to observations that Sardine eggs had become more common south of Kangaroo Island (e.g. Ward et al. 2018b, 2019) and that commercial catches were increasing in the area south-east of Kangaroo Island (Grammer et al. 2021). The widespread occurrence of Sardine eggs at the new sampling sites continues to provide evidence that the expansion of the survey area was warranted and should be maintained in future years.

4.2. Spawning area and mean daily egg production

The spawning area in 2022 was estimated to be 76,842 km², which is the second largest on record. The largest spawning area was recorded in 2020 at 82,627 km². The high spawning area of 2022 includes the additional survey area added to the east of Kangaroo Island. Excluding the additional sites still results in a spawning area of 70,576 km², which is the third highest on record (73,981 km² recorded in 2014) for South Australian Sardine.

Spawning area is strongly correlated with Sardine abundance (Mangel and Smith 1990, Gaughan et al. 2004), and Ward et al. (2021) confirmed that spawning area is a good proxy for the abundance of adult Sardine off South Australia. The large spawning area observed in this study provides evidence that Sardines were widespread and abundant off South Australia in 2022. Using historical data to estimate all DEPM parameters except spawning area means that fluctuations in estimates of spawning biomass are driven entirely by changes in the measure of spawning area. As a result, future surveys must cover as much of the spawning area as possible and should continue to involve the adaptive approach to sampling that has been in place from 2014 onwards.

Inter-annual variability in estimates of P_0 is low compared to statistical uncertainty (imprecision) for Sardine off South Australia (Ward et al. 2021). The estimate of P_0 obtained by adding the 2022 data to all historical data was more precise (SD = 4.4) than the estimate obtained using data from 2022 only (SD = 19.0). Using historical data to estimate of P_0 prevents large inter-annual fluctuations in estimates of spawning biomass driven by variations

in the annual estimates of this parameter caused by statistical uncertainty. However, it is critical that P_0 continue to be monitored at a yearly timescale to look for changes in the parameter over time to reduce the risk of over- or under-estimating spawning biomass (e.g. the basin hypothesis of McCall 1990) (Ward et al. 2021). In future applications of the DEPM to Sardine off South Australia, P_0 should continue to be estimated using data obtained in all years since 1998, as well as within each year.

4.3. Adult parameters

Re-analysis of adult samples of Sardine collected off South Australia since 1998 suggest that both individual parameters and mean daily fecundity are relatively stable among years, especially when inter-annual variability is evaluated within the context of potential sources of statistical uncertainty (i.e. precision and bias) (Ward et al. 2021). Large variations among years observed in the estimates of the adult parameters are more likely to reflect the limitations of the adult sampling program, rather than actual differences among years in the reproductive patterns of the population (Ward et al. 2021). Therefore, in the foreseeable future, adult parameters used to calculate the spawning biomass of Sardine off South Australia should be estimated from data obtained in adult surveys conducted between 1998 and 2018.

While no adult parameter was correlated with spawning area (Ward et al. 2021)—correlation would suggest density dependent changes had occurred—detecting changes in the adult population over time due to environmental conditions or density dependent effects is needed. Adult sampling is costly and logistically challenging, yet necessary. In recent years, completing a successful ichthyoplankton survey that covers the entire spawning area has taken precedence over adult sampling. Detecting any changes in key adult parameters of Sardine, such as spawning fraction (*S*), is critical since errors when estimating these parameters have major implications for estimates of spawning biomass (Ward et al. 2021). To mitigate this risk, an extensive adult sampling program needs to be conducted within the next few years and repeated periodically (e.g. every 3-5 years) to determine if changes in key adult parameters have have occurred.

4.4. Spawning biomass

The estimate of spawning biomass for 2022 of 355,075 (299,956–410,193) t is above the upper target reference point of 190,000 t in the harvest strategy for the South Australian Sardine Fishery (PIRSA 2014). On this basis, the southern stock of Australian Sardine is classified as **Sustainable**. This classification is consistent with recent assessments provided in the spawning biomass report for 2020 (Ward et al. 2020), the stock assessment report for

23

2021 (Grammer et al. 2021) and the most recent report on the Status of Australian Fish Stocks (Piddocke et al. 2021).

REFERENCES

- Brooks, M. E., K. Kristensen, K. J. v. Benthem, A. Magnusson, C. W. Berg, A. Nielsen, and B. M. Bolker. 2017. glmmTMB Balances Speed and Flexibility Among Packages for Zero-inflated Generalized Linear Mixed Modeling. The R Journal 9:378-400.
- Dimmlich, W. F., T. M. Ward, and Breed, W.G. 2009. Spawning dynamics and biomass estimates of an anchovy *Engraulis australis* population in contrasting gulf and shelf environments. Journal of Fish Biology **75**:1560–1576.
- Fletcher, W. J., N. C. H. Lo, E. A. Hayes, R. J. Tregonning, and S. J. Blight. 1996. Use of the daily egg production method to estimate the stock size of western Australian sardines (*Sardinops sagax*). Marine and Freshwater Research **47**:819–825.
- Gaughan, D. J., T. I. Leary, R. W. Mitchell, and I. W. Wright. 2004. A sudden collapse in distribution of Pacific sardine (*Sardinops sagax*) off southwestern Australia enables an objective reassessment of biomass estimates. Fishery Bulletin **102**:617–633.
- Grammer, G., F. Bailleul, A. Ivey, and J. Smart. 2021. Stock assessment of Australian Sardine (*Sardinops sagax*) off South Australia 2021. Report to PIRSA Fisheries and Aquaculture. South Australian Research and Development Institute (Aquatic Sciences), Adelaide. SARDI Publication No. F2007/000765-8. SARDI Research Report Series No. 1120. 96pp.
- Grammer, G. L., Ward, T.M. and Durante, L.M. 2022. Commonwealth Small Pelagic Fishery: Fishery Assessment Report 2019-2021. Report to the Australian Fisheries Management Authority. South Australian Research and Development Institute (Aquatic Sciences), Adelaide. SARDI Publication No. F2010/000270-11. SARDI Research Report Series No. 1133. 101pp.
- Hunter, J. R., and S. R. Goldberg. 1980. Spawning incidence and batch fecundity in northern anchovy, Engraulis mordax. FIsheries Bulletin **77**:641–652.
- Hunter, J. R., and B. J. Macewicz. 1985. Measurement of spawning frequency in multiple spawning fishes. Pages 79–94 *in* R. Lasker, (eds). An egg production method for estimating spawning biomass of pelagic fish: Application to the northern anchovy, Engraulis mordax., NOAA Tech. Rep. NMFS.
- Lasker, R. 1985. An egg production method for estimating spawning biomass of pelagic fish: application to northern anchovy, *Engraulis mordax*. NOAA. Tech. Rep. NMFS **36**:1–99.
- Le Clus, F., and P. Malan. 1995. Models of temperature-dependent rate of development of pilchard *Sardinops sagax* eggs, to be used in routine procedures for estimating daily egg production. South African Journal of Marine Science 16:1-8.
- Mangel, M., and Smith, P.E. (1990) Presence-absence sampling for fisheries management. Canadian Journal of Fisheries and Aquatic Sciences **47**: 1875–1887.
- McCall, A. D. 1990. Dynamic Geography of Marine Fish Populations. University of Washington Press, Seattle, WA.
- Neira, F. J., A. G. Miskiewicz, and T. Trnski. 1998. Larvae of temperate Australian fishes: Laboratory guide for larval fish identification. University of Western Australia Press: Western Australia.
- Neira, F. J., J. M. Lyle, and J. P. Keane. 2009. Shelf spawning habitat of *Emmelichthys nitidus* in south-east Australia - Implications and suitability for egg-based biomass estimation. Estuarine Coastal and Shelf Science 81:521-532.
- Parker, K. 1980. A direct method for estimating northern anchovy, *Engraulis mordax*, spawning biomass. . Flsheries Bulletin **78**:541–544.
- Pauly, D., and R. S. V. Pullin. 1988. Hatching time in spherical, pelagic, marine fish eggs in response to temperature and egg size. Environmental biology of fishes 22:261-271.
- Picquelle, S., and G. Stauffer. 1985. Parameter estimation for an egg production method of anchovy biomass assessment. Pages 7–17 *in* R. Lasker, (eds). An egg production method for estimating spawning biomass of pelagic fish: application to northern anchovy, *Engraulis mordax*. NOAA Tech. Rep. NMFS.
- PIRSA. 2014. Management plan for the South Australian commercial marine scalefish fishery. Part B–Management arrangements for the taking of Sardines. Paper number 68.
- R Core Team. 2022. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL http://www.R-project.org/.
- Smith, P. E., and S. L. Richardson. 1977. Standard techniques for pelagic fish egg and larva surveys. FAO Fisheries Technical Paper **175**:100p.

- Somarakis, S., I. Palomera, A. Garcia, L. Quintanilla, C. Kousikopoulos, A. Uriarte, and L. Motos. 2004. Daily egg production of Anchovy in European waters. ICES Journal of Marine Science. **61**.
- Stratoudakis, Y., M. Bernal, K. Ganias, and A. Uriate. 2006. The daily egg production method: recent advances, current applications and future challenges. Fish and Fisheries 7:35–57.
- Piddocke, T., Ashby, C., Hartmann, K., Hesp, A., Hone, P., Klemke, J., Mayfield, S., Roelofs, A., Saunders, T., Stewart, J., Wise, B. and Woodhams, J. (eds) 2021. Status of Australian fish stocks reports 2020, Fisheries Research and Development Corporation, Canberra.
- Turner, R. 2016. deldir: Delaunay Triangulation and Dirichlet (Voronoi) Tessellation. R package version 0.1–12. https://CRAN.R-project.org/package=deldir
- Ward, T. M., M. Kinloch, J. G.K., and F. J. Neira. 1998. A Collaborative Investigation of the Usage and Stock Assessment of Baitfish in Southern and Eastern Australian Waters, with Special Reference to Pilchards (*Sardinops sagax*). South Australian Research and Development Institute (Aquatic Sciences). Final Report to Fisheries Research and Development Corporation.
- Ward, T. M., P. Burch, L. J. McLeay, and A. R. Ivey. 2011. Use of the Daily Egg Production Method for stock assessment of sardine, *Sardinops sagax*; lessons learned over a decade of application off southern Australia Reviews in Fisheries Science **19**:1–20.
- Ward, T. M., J. Carroll, G. L. Grammer, C. James, R. McGarvey, J. Smart, and A. R. Ivey. 2018a. Improving the precision of estimates of egg production and spawning biomass obtained using the Daily Egg Production Method. South Australian Research and Development Institute (Aquatic Sciences), Adelaide. FRDC project No. 2014/026. 90 pp.
- Ward, T. M., G. L. Grammer, A. R. Ivey, J. J. Smart, and J. P. Keane. 2018b. Spawning biomass of Jack Mackerel (*Trachurus declivis*) and Sardine (*Sardinops sagax*) between western Kangaroo Island, South Australia and south-western Tasmania. Report to the Australian Fisheries Management Authority. SARDI Aquatic Sciences, Adelaide. SARDI Publication No. F2018/000174-1. SARDI Research Report Series No. 983. 51pp.
- Ward, T.M., Ivey, A.R. and Grammer, G.L. 2019. Spawning biomass of Sardine, Sardinops sagax, in waters off South Australia in 2019. Report to PIRSA Fisheries and Aquaculture. South Australian Research and Development Institute (Aquatic Sciences), Adelaide. SARDI Publication No. F2007/000566-9. SARDI Research Report Series No. 996. 31pp.
- Ward, T.M., Ivey, A.R. and Grammer, G.L. 2020. Spawning biomass of Sardine, Sardinops sagax, in waters off South Australia in 2020. Report to PIRSA Fisheries and Aquaculture. South Australian Research and Development Institute (Aquatic Sciences), Adelaide. SARDI Publication No. F2007/000566-11. SARDI Research Report Series No. 1074. 28pp.
- Ward, T.M., Grammer, G.L., Ivey, A., Smart, J.J. and McGarvey, R. 2021. Increasing the precision of the daily egg production method; 2020's remix of a 1980's classic. ICES Journal of Marine Science 78:1177–1195.
- Watson, D. F. 1981. Computing the n-dimensional Delaunay tessellation with application to Voronoi polytopes. Pages 167-172 The Computer Journal.
- White, K. V., and W. J. Fletcher. 1998. Identifying the developmental stages for eggs of the Australian pilchard, *Sardinops sagax*. Fisheries Western Australia. Fisheries Research
- Wood, S. N. 2006. Generalized Additive Models An Introduction with R. Chapman and Hall/CRC.Report No.103.

APPENDIX 1: ANNUAL AND ALL-YEARS PARAMETERS FOR ESTIMATES OF SPAWNING BIOMASS

Annual and all-years parameters used to calculate estimates of Spawning Biomass. Total *A*: total area sampled (km²), *A*: spawning area (km²); *P*₀: mean daily egg production (egg·m⁻²·day⁻¹); *S*: spawning fraction; *R*: sex ratio; *W*: mean female weight (g); \hat{F} : batch fecundity (oocytes·batch⁻¹); *F*': Fecundity / Female Weight. Errors around the estimates are standard deviation (SD). N: number of samples; n: number of individuals. *F*' was calculated using the all-years \hat{F} relationship with *W* from that year. Data sources for table: Ward et al. (2021) and Grammer et al. (2021).

Time	Total A	Α	P ₀	P ₀	N.P₀	S	S	N.S	n.S	R	R	N.R	n.R	W	W	N.W	n.W	Ê	Ê	N. Ê	n. Ê	F'	F'
				SD			SD				SD				SD				SD				SD
All Years	-	-	83.6	4.4	6432	0.108	0.006	247	16334	0.55	0.01	210	27931	58.4	18.0	255	16995	17835	7166	255	16995	305.4	0.6
1998	48379	32980	99.0	30.8	164	0.139	0.015	12	530	-	-	-	-	46.5	11.2	12	554	14070	5295	12	554	302.3	3.8
1999	65956	15637	50.0	14.9	213	0.169	0.021	15	763	-	-	-	-	52.4	13.0	16	785	15743	5616	16	785	300.7	3.0
2000	102198	38658	52.9	12.7	290	0.158	0.012	15	1012	0.52	0.05	15	2179	49.2	12.2	16	1071	14543	5416	16	1071	295.9	2.5
2001	132382	39131	59.7	15.6	316	0.179	0.014	10	743	0.56	0.04	10	1397	50.7	9.1	11	1002	15614	5069	11	1002	307.7	2.7
2002	131574	37462	97.4	29.1	319	0.077	0.014	22	1631	0.60	0.04	22	2932	61.8	19.5	22	1841	19093	7664	22	1841	309.0	1.8
2003	133058	42905	113.5	27.4	320	0.103	0.008	7	435	0.48	0.03	7	986	52.4	8.5	7	435	16179	4720	7	435	308.9	3.9
2004	105621	40219	145.3	41.3	284	0.166	0.016	10	412	0.52	0.04	10	879	78.7	16.2	10	413	23995	7607	10	413	304.8	3.7
2005	122831	42142	59.5	14.3	334	0.100	0.019	32	2223	0.51	0.04	32	4827	73.9	16.0	33	2234	22426	7367	33	2234	303.5	1.6
2006	119038	50121	102.4	26.5	341	0.095	0.018	20	1332	0.59	0.05	20	2445	63.1	21.8	21	1337	19163	8194	21	1337	303.6	2.2
2007	119036	50972	104.9	27.1	341	0.130	0.019	20	1084	0.54	0.07	20	2244	71.1	16.8	21	1086	21723	7266	21	1086	305.7	2.3
2009	119031	55179	66.3	14.1	340	0.156	0.022	19	1537	0.36	0.04	9	2425	59.9	13.3	19	1537	17886	6183	19	1537	298.7	2.0
2011	119449	44245	51.5	15.4	340	0.044	0.006	14	1169	0.65	0.05	13	1798	46.8	12.3	15	1181	14640	5487	15	1181	312.9	2.5
2013	119297	37953	39.0	8.7	340	0.072	0.016	9	703	0.69	0.03	9	1089	51.3	12.3	9	723	15842	5761	9	723	309.1	3.2
2014	125249	73981	92.7	20.0	355	0.041	0.006	16	886	0.57	0.02	16	1574	47.9	13.9	16	886	14896	5983	16	886	311.1	3.0
2016	122598	50551	47.7	9.9	350	0.088	0.012	9	656	0.65	0.03	9	1088	49.7	17.3	9	681	15399	6774	9	681	309.6	3.3
2017	119661	66453	136.3	25.8	343	0.120	0.019	8	504	0.52	0.05	9	1042	59.6	12.3	9	511	18331	6250	9	511	307.8	3.7
2018	120043	63215	112.4	24.6	343	0.054	0.009	9	714	0.70	0.03	9	1026	46.5	7.2	9	718	14466	4430	9	718	311.0	3.0
2019	119369	53600	68.1	16.3	339	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
2020	129700	82627	94.0	18.4	379	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
2022	129982	76842	97.6	19.0	381																		